

How Do Atrial-Selective Drugs Differ From Antiarrhythmic Drugs Currently Used in the Treatment of Atrial Fibrillation?

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Abstract

Current pharmacologic strategies for the management of atrial fibrillation (AF) include use of 1) sodium channel blockers, which are contraindicated in patients with coronary artery or structural heart disease because of their potent effect to slow conduction in the ventricles, 2) potassium channel blockers, which predispose to acquired long QT and Torsade de Pointes arrhythmias because of their potent effect to prolong ventricular repolarization, and 3) mixed ion channel blockers such as amiodarone, which are associated with multi-organ toxicity. Accordingly, recent studies have focused on agents that selectively affect the atria but not the ventricles. Several atrial-selective approaches have been proposed for the management of AF, including inhibition of the atrial-specific ultrarapid delayed rectified potassium current (IKur), acetylcholine-regulated inward rectifying potassium current (IK-ACh), or connexin-40 (Cx40). All three are largely exclusive to atria. Recent studies have proposed that an atrial-selective depression of sodium channel-dependent parameters with agents such as ranolazine may be an alternative approach capable of effectively suppressing AF without increasing susceptibility to ventricular arrhythmias. Clinical evidence for Cx40 modulation or IK-ACh inhibition are lacking at this time. The available data suggest that atrial-selective approaches involving a combination of INa, IKur, IKr, and, perhaps, Ito block may be more effective in the management of AF than pure IKur or INa block. The anti-AF efficacy of the atrial-selective/predominant agents appears to be similar to that of conventionally used anti-AF agents, with the major apparent difference being that the latter are associated with ventricular arrhythmogenesis and extracardiac toxicity.

Introduction

The development of atrial-selective antiarrhythmic agents was necessitated by the proclivity for induction of life-threatening ventricular arrhythmias and/or extra-cardiac toxicity of currently available anti-atrial fibrillation (AF) agents. This review is an attempt to briefly summarize currently current

knowledge relative to this effort.

It has been more than a decade since Wang et al first described the ultrarapid delayed rectified potassium current (IKur, carried by Kv 1.5 channels encoded by the KCNA5 gene) which is present only in atria, but not ventricles. Block of IKur affects only atrial electrical parameters, which

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makes this current a potential target for chamber-specific management of atrial arrhythmias.² Among other atrial specific targets suggested to be used for the suppression of atrial arrhythmias is the acetylcholine (ACh)-regulated inward rectifying potassium current (IK-ACh), encoded by Kir3.1/3.4 alpha-subunits) and, to a certain degree, connexin 40 (Cx40, encoded by GJA5 and).^{2,4}

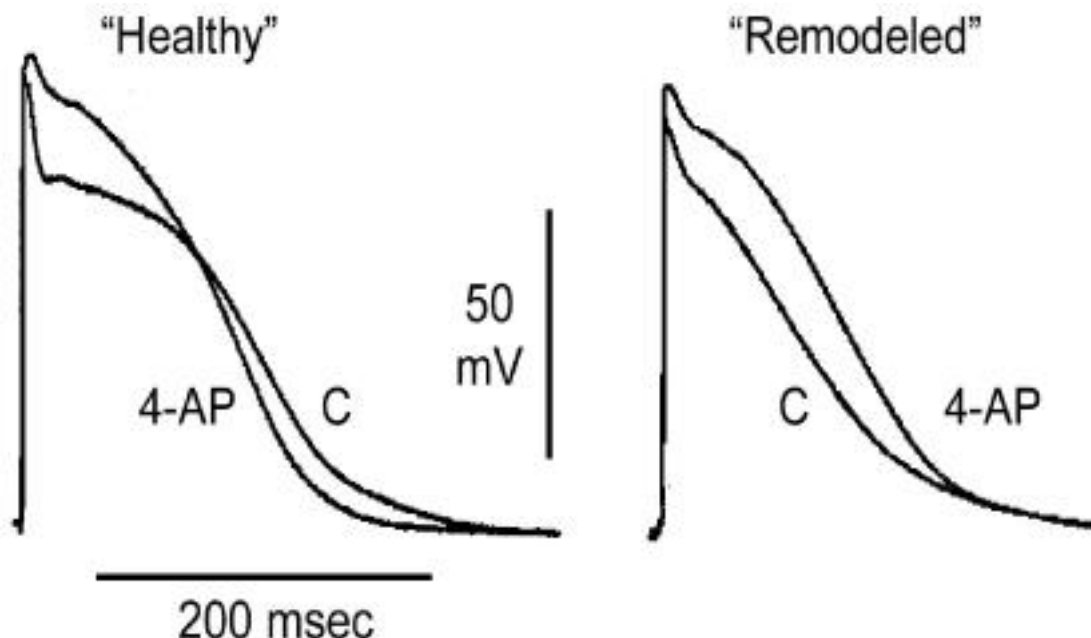
Most recently, another approach has been proposed for the management of AF, consisting of atrial-selective/predominant depression of sodium channel current (INa)-dependent parameters. INa blockers, such as ranolazine, which predominantly or selectively, depresses INa-dependent parameters in atria vs. ventricles.⁵ Ranolazine has been shown to be effective in suppressing AF in a variety of experimental models⁵ as well as reducing supraventricular arrhythmias ($p < 0.001$) and new onset AF ($p = 0.08$) in patients with non-ST segment elevation acute coronary syndrome.⁶ Block of IKr has also been shown to cause a preferential prolongation of atrial vs. ventricular repolarization.⁷⁻¹³

Atrial-Specific Antiarrhythmic Approaches for AF

Atrial-specific targets are those that are present exclusively or almost exclusively in atria. Atrial-specific agents include those that inhibit IKur and IK(ACh). Agents that modulate Cx40, found in atrial but not ventricular myocardium, are commonly included in this category with the caveat that Cx40 is present in the conduction system of the ventricles.^{2,4} While it is conceivable that an effect on Cx40 may suppress some forms of AF, there are no specific Cx40 inhibitors available yet, so that the hypothesis remains to be tested.

Vagally-mediated AF is the most likely form of AF to be suppressed by IK-ACh inhibition. A vagal component may also contribute to the initiation of paroxysmal AF.^{14,15} Normally, IK-ACh is activated through the muscarinic receptors in response to release of ACh, leading to an abbreviation of atrial repolarization and, thereby, promoting AF. In the atria of human patients with chronic AF, ACh-activated IK-ACh is reported to be either increased¹⁶ or decreased.³ There is another form of IK-ACh that

Figure 1: Block of IKur with 4-aminopyridine (4-AP, 50 μ M)



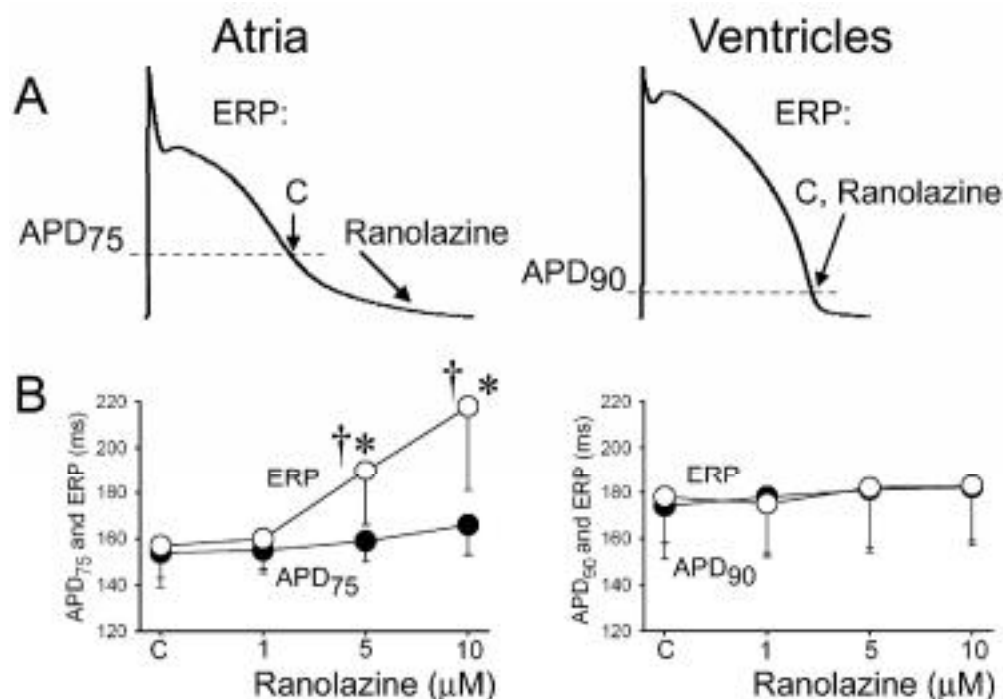
abbreviates APD90 in "healthy" (plateau-shaped action potential), but prolongs it in "acutely remodeled" (triangular-shaped action potential) canine coronary-per fused Atrial preparations (pectinate muscles). Low flow ischemia was used to generate the "acutely remodeled" atria. Left panel is from Burashnikov et al.,²⁴ with permission.

does not require vagal influences or muscarinic receptors for activation, i.e., the constitutively active IK-ACh.¹⁷⁻¹⁹ This current is present in normal human atria and is significantly increased in atria of chronic AF patients.^{18, 19} In canine atria, a corresponding constitutively active IK-ACh is also present under baseline conditions and this current is up-regulated in tachycardia-remodeled atria.^{17, 20} The constitutively active IK-ACh is likely to contribute to abbreviation of atrial APD and AF vulnerability.¹⁸⁻²⁰ Selective block of IK-ACh with tertiapin prolongs atrial APD and suppresses AF in experimental AF models.^{20, 21} Block of IK-ACh, however, may be pro-arrhythmic in post-operative AF cases, since the preservation of the anterior fat pad (containing mostly parasympathetic nerves endings) decreases the incidence of postoperative AF.²² There are no clinically available selective IK-ACh blockers at the present time. An important caveat in the development of clinically safe IK-ACh blockers is that these agents should not produce significant vagolytic influences in other organs. Thus, the

clinical feasibility of an "IKA-Ch approach" is yet to be established.

Among atrial-specific targets, IK_{ur} is the most investigated and is considered by many as the most promising target at the present time. The pharmacological response of "healthy" and "remodeled" atria (displaying a plateau-shaped and triangular-shaped AP morphology, respectively) to IK_{ur} block is very different (Fig. 1).^{23, 24} Block of IK_{ur} in "healthy" atria prolongs only the early repolarization phase but abbreviates the late repolarization phase (APD₇₀ or APD₉₀) and the effective refractory period (ERP)²³⁻²⁵ In contrast, in remodeled atria, a reduction of IK_{ur} prolongs APD₇₀₋₉₀.²³ Interestingly, loss-of-function mutations in KCNA5, the gene that encodes the Kv1.5 channel protein, have been associated with familial AF,²⁶ suggesting that a reduction in IK_{ur} may predispose to the development of AF. In support of this hypothesis, block of IK_{ur} with 10-50 μ M of 4-AP was shown to promote the induction of non-sustained AF in "healthy" canine arterially-perfused atrial prepa-

Figure 2: Ranolazine specifically induces prolongation of the effective refractory period (ERP) and development of post-repolarization refractoriness in atria



(PRR, the difference between ERP and APD₇₅ in atria and between ERP and APD₉₀ in ventricles; ERP corresponds to APD₇₅ in atria and APD₉₀ in ventricles). CL = 500 ms. C – control. The arrows in panel A illustrate the position on the action potential corresponding to the end of the ERP in atria and ventricles and the effect of ranolazine to shift the end of the ERP in atria but not ventricles. * $p < 0.05$ vs. control. † $p < 0.05$ vs. APD₇₅ values in atria and APD₉₀ in ventricles; (n=5-18). From Burashnikov et al⁵ with permission.

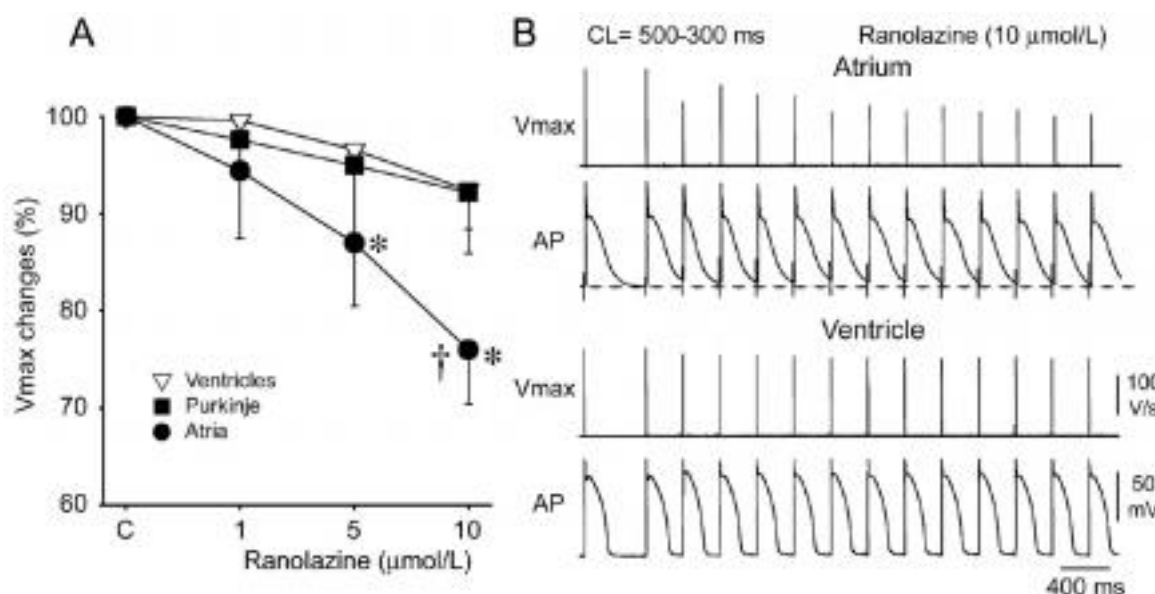
rations, secondary to the development of an abbreviation of APD90 and ERP.²⁵

A number of studies have demonstrated that agents capable of blocking I_{Kur} (AVE0118, AVE1231, S9947, S20951, ISQ-1, DPO-1, vernakalant; AZD7009; NIP141, NIP-142) selectively prolong atrial ERP both in electrically-remodeled and non-remodeled (i.e., "healthy") atria in vivo and in vitro.²⁷⁻³⁴ Because ERP prolongation in "healthy" atria is not consistent with APD70-90 abbreviation recorded in "healthy" atrial preparations in vitro,²³⁻²⁵ the ERP prolongation observed in response to these agents may be related to their action to also depress I_{Na} . Prolongation of ERP in the absence of APD90 prolongation is a well-known feature of sodium, but not potassium, channel blockade. This is the result of post-repolarization refractoriness (PRR), which develops more readily in atria than in the ventricles (Fig. 2).^{5, 35, 36} Consistent with this line of thinking, I_{Kur} blockers such as AZD7009 and vernakalant have been also shown to be capable of potentially block I_{Na} .^{32, 37, 38} AZD7009 has been shown to behave as an atrial selective I_{Na} blocker in the canine heart in vivo, slowing conduction velocity and increasing diastolic threshold of excitation in atria, but not ventricles.³² ISQ-1

may also block I_{Na} , since this agent slows down conduction velocity in atria in vivo.³⁹ AVE0118 prolongs ERP but not APD90 and reduces V_{Max} in canine coronary-perfused atrial preparations (Burashnikov et al, unpublished). Note that with the exception of AZD7009 (see above), comparison of the effects of I_{Kur} blockers on I_{Na} or sodium channel-dependent parameters in atria and ventricles has not been conducted.

I_{Kur} blockers that have been shown to be experimentally and clinically effective in terminating AF (i.e., AZD7009 and vernakalant) have also proven to exert potent block of I_{Na} and I_{Kr} .^{37, 38} It is not clear whether I_{Kur} or I_{Na} plays the greater role in producing atrial-selective prolongation of ERP and suppression of AF. Perhaps the most-investigated I_{Kur} blocker, AVE0118, suppresses AF in goats and pigs, but at concentrations that inhibit Ito and I_{K-ACh} .^{27, 40} Another I_{Kur} blocker ISQ-1 can terminate AF in in vivo dogs at the concentrations that block I_{Kr} and possibly I_{Na} , since conduction time is significantly increased in atria.³⁹ Low concentrations of 4-AP (10-50 μ M, which selectively inhibit I_{Kur}) do not prevent the initiation of AF or terminate persistent ACh-mediated AF in canine coronary-perfused atrial

Figure 3: Ranolazine produces a much greater rate-dependent inhibition of the maximal action potential upstroke velocity (V_{max}) in atria than in ventricles.



A: Normalized changes in V_{max} of Atrial and ventricular cardiac preparations paced at a cycle length (CL) of 500 ms. C: Ranolazine prolongs late repolarization in atria, but not ventricles and acceleration of rate leads to elimination of the diastolic interval (during which the recovery from sodium channel block occurs), resulting in a more positive take-off potential in atrium and contributing to Atrial selectivity of ranolazine. The diastolic interval remains relatively long in ventricles. * $p < 0.05$ vs. control. † $p < 0.0$ vs. respective values of M cell and Purkinje ($n = 7-21$). From Burashnikov et al with permission.

preparations.²⁵

Thus, available experimental and clinical data suggest that “pure” IKur inhibition alone is incapable of effectively suppressing AF and that the antiarrhythmic effects of most IKur blockers under development are attributable to associated inhibition of other ion channels including INa, Ito, and/or IKr). The relative contribution of IKur reduction is not clear at this time.

Atrial Predominant Antiarrhythmic Approaches for AF.

We refer to atrial selective or predominant targets as to those that are present in both chambers of the heart, but inhibition of these targets produces greater effects in atria vs. ventricles. “Atrial-predominant” refers to a lesser degree of atrial selectivity. The results of recent experimental studies indicate that some INa blockers (such as ranolazine and chronic amiodarone) depress sodium channel-dependent parameters in an atrial selective or predominant manner.^{5, 35, 36} While not well appreciated, IKr blockers are also atrial-predominant in that they preferentially prolong atrial repolarization at normal heart rates.⁷⁻¹³

Atrial Predominant Effects of IKr Block on Cardiac Repolarization

In direct comparisons, selective IKr blockers such as E-4031, sotalol, d-sotalol, dl-sotalol, dofetilide, WAY-123,398, ibutilide, MK499, and almokalant preferentially prolong atrial vs. ventricular ERP and APD at normal pacing rates.⁷⁻¹³ At slow pacing/heart rates, however, ventricles, but not atria commonly develop exaggerated APD prolongation, early afterdepolarizations, and Torsade de Pointes in response to a reduction of IKr.^{41, 42}

Does Ito Block Preferentially Alter Atrial vs. Ventricular Repolarization?

The IC₅₀ of 4-AP’s action to block atrial Ito is one third of that of ventricular Ito in human and canine myocytes.^{43, 44} If this proves to be the case with other Ito blockers, then Ito block might be expected to produce a greater effect on atrial vs. ventricular repolarization. Because most IKur blockers also reduced Ito, inhibition of Ito may

contribute to the atrial-selectivity of IKur blockers, both with respect to their antiarrhythmic and proarrhythmic actions.

Atrial Predominant Sodium Channel Block

In recent studies, we examined atrioventricular differences of the effects of ranolazine, chronic amiodarone, lidocaine and propafenone on sodium channel-dependent parameters, such as the maximum rate of rise of the action potential upstroke (V_{max}), diastolic threshold of excitation (DTE), conduction velocity (CV), and PRR.^{5, 35, 36, 45} Using canine isolated coronary-perfused atrial and ventricular preparations, we evaluated therapeutically-relevant concentrations of these agents. Ranolazine, a recently marketed antianginal agent, was found to depress V_{max}, DTE, CV, and induce PRR exclusively or predominantly in atrial preparations.⁵ Thus, when studied in beating multicellular preparations, ranolazine proved to be an atrial-selective/predominant sodium channel blocker (“an atrial selective Class I agent”).

Chronic amiodarone was found to depress sodium-channel dependent parameters in both atrial and ventricular preparations, but much more effectively in atria.^{35, 36} Lidocaine was also atrial-predominant, but far less atrial-selective than ranolazine or amiodarone.⁵ Propafenone showed no chamber selectivity for INa block at a normal pacing rate (CL = 500 ms), but some atrial predominance at rapid pacing rates, likely due to atrial specific APD prolongation (see later).⁴⁵ As previously mentioned, AZD7009 also behaves as an atrial-selective INa blocker.³²

Ranolazine, propafenone, and chronic amiodarone all produce prolongation of APD₉₀ in the canine atria, likely to due to their actions to inhibit IKr. This effect of these agents potentiate their effects to reduce INa and depress INa-dependent parameters, thus contributing to their atrial-selective effects, particularly at rapid activation rates (Fig. 3C).

The atrial-selective action of these agents is thought to be due to important distinctions in action potential characteristics as well as biophysical properties of sodium channels of atrial and

ventricular myocytes. The half inactivation voltage ($V_{0.5}$) in canine atrial myocytes is 12-16 mV more negative than that of ventricular myocytes and resting membrane potential (RMP) in atria is less negative than ventricles (approx. -83 vs. -87 mV).^{5,46} These factors indicate that there is a larger fraction of inactivated sodium channels at RMP in atria vs. ventricles and a smaller fraction of resting sodium channels at RMP in atria vs. ventricles. This is expected to slow the unbinding of the drug and recovery of the sodium channel from pharmacologic block in atria, since this recovery occurs principally during the resting state.⁴⁷

Anti-AF Potential of Atrial Selective vs. Conventional Agents

We compared the effectiveness of therapeutically-relevant concentrations of ranolazine, propafenone, and lidocaine in suppressing and preventing the re-induction of AF in isolated canine coronary-perfused right atrial preparations.^{5, 35, 45} The effectiveness of chronic amiodarone in preventing induction of AF was examined as well. Ranolazine prevented the initiation acetylcholine-mediated AF, terminated persistent AF, and prevented its re-induction in coronary-perfused atrial preparations (Fig. 6).⁵ This anti-AF efficacy of ranolazine (10 μ M) was greater than that of lidocaine (21 μ M) and similar to that of propafenone (1.5 μ M). In atria isolated from chronic amiodarone-treated dogs (40 mg/Kg for 6 weeks), persistent ACh-mediated AF could be induced only in 1 out of 6 atria (vs. 10/10 atria isolated from untreated controls).³⁵ These antiarrhythmic effects of ranolazine, amiodarone, and propafenone were associated with both APD prolongation (in the presence of ACh) and the development a significant PRR, with the duration of the latter being much longer the extent of APD prolongation, suggesting that sodium-channel block plays a more predominant role in the anti-AF actions of these agents.^{5,35}

Ranolazine and propafenone both suppress AF but ranolazine, unlike propafenone, does it without prominent effects on ventricular myocardium. These findings suggest that atrial-selective/predominant sodium channel block, perhaps with additional IKr block, may be a promising new atrial-selective approach for the management of AF.⁵ Interestingly, all clinically effective anti-AF INa

blockers also inhibit IKr. Pure INa blockers, such as lidocaine, are not very effective in suppression of AF in the clinic.

Clinical efficacy has been reported for only three atrial-selective agents: AZD7009, vernakalant (agents that block IK_{ur}, INa, and IKr) and ranolazine, which blocks INa and IKr. There is therefore little basis for a comparison of atrial-selective agents with conventional anti-AF agents. In a study that was not designed to test the anti-AF efficacy of ranolazine, this agent was found to reduce the incidence of new onset AF onset by 30% in acute coronary syndrome patients ($p=0.08$).⁶ AZD7009 was reported to successfully convert up to 70% of patients with an average AF duration of 43 days to sinus rhythm⁴⁸ whereas vernakalant was successful in converting 52-56% of patients with recent AF onset (< 7 days) but only 8% of patients with long-duration AF (< 45 days).⁴⁹ These findings are not very different from those of conventionally used anti-AF agents, such as flecainide, propafenone, dofetilide, amiodarone, ibutilide, etc.⁵⁰ The benefit of the atrial selective agents is that they do not produce significant electrophysiological changes in the ventricles. It appears that amiodarone can also be categorized as atrial-selective.³⁵ It is noteworthy that direct comparisons of the effects of most clinically-used anti-AF agents in atria and ventricles are not available either in *in vivo* or in coronary-perfused preparation studies.

Current pharmacologic strategies for the management of AF include sodium channel blockers such as propafenone and flecainide, potassium channel blockers such as sotalol and dofetilide and mixed ion channel blockers such as amiodarone. All have demonstrated efficacy but distinct indications based on their proclivity to promote ventricular arrhythmogenesis under different conditions.⁵⁰ These adverse effects distinguish these agents from the newer atrial-selective agents. The sodium channel blockers are contraindicated in patients with coronary artery or structural heart disease because of their potent effect on conduction in the ventricles, potassium channel blockers predispose to the development of acquired long QT and Torsade de Pointe arrhythmias because of their potent effect to prolong ventricular repolarization, and mixed ion channel blockers such as

amiodarone are associated with multi-organ toxicity.

Atrial Selectivity and Atrial Remodeling.

It is important to recognize that the atrial selective/predominant effects of some INa as well as IKr blockers have been tested in relatively “healthy” atria and ventricles.^{5, 7-10, 35, 35} Clinical AF commonly occurs in conjunction with a number of conditions (congestive heart failure, hypertrophy, dilatation, hypertension, etc) associated with electrical and/or structural remodeling of the atria. These pathophysiological changes can importantly modify the response to sodium and potassium channel blockers,^{23, 51, 52} and thus modify the atrial selectivity of these agents.

Conclusions

There is no robust evidence in support of the hypothesis that “pure” inhibition of IKur may effectively suppress AF. No clinical data are available relative to the anti-AF efficacy and safety of IKACH or Cx40 as atrial-specific approaches. The available data suggest that atrial-selective approaches involving a combination of INa, IKur, IKr, and, perhaps Ito blockade, may be more effective in the management of AF than pure IKur or INa block. The anti-AF efficacy of the atrial-selective/predominant agents appears to be similar to that of conventionally used anti-AF agents, with the major difference being that the latter are associated with ventricular arrhythmogenesis as well as extracardiac toxicity. It is noteworthy, however, that the long-term toxicity of the atrial-selective drugs, with the exception of ranolazine, is not known.

Disclosures

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References

1. Wang ZG, Fermini B, Nattel S. Sustained depolarization-induced

outward current in human atrial myocytes: Evidence for a novel delayed rectifier K⁺ current similar to Kv1.5 cloned channel currents. *Circ Res.* 1993;73:1061-1076.

2. Nattel S, Carlsson L. Innovative approaches to anti-arrhythmic drug therapy. *Nat Rev Drug Discov.* 2006;5:1034-1049.

3. Dobrev D, Graf E, Wettwer E, Himmel HM, Hala O, Doerfel C, Christ T, Schuler S, Ravens U. Molecular basis of down-regulation of G-protein-coupled inward rectifying K(+) current (I(K,ACh) in chronic human atrial fibrillation: decrease in GIRK4 mRNA correlates with reduced I(K,ACh) and muscarinic receptor-mediated shortening of action potentials. *Circulation.* 2001;104:2551-2557.

4. Ehrlich JR, Biliczki P, Hohnloser SH, Nattel S. Atrial-selective approaches for the treatment of atrial fibrillation. *J Am Coll Cardiol.* 2008;51:787-792.

5. Burashnikov A, Di Diego JM, Zygmunt AC, Belardinelli L, Antzelevitch C. Atrium-selective sodium channel block as a strategy for suppression of atrial fibrillation: differences in sodium channel inactivation between atria and ventricles and the role of ranolazine. *Circulation.* 2007;116:1449-1457.

6. Scirica BM, Morrow DA, Hod H, Murphy SA, Belardinelli L, Hedgepeth CM, Molhoek P, Verheugt FW, Gersh BJ, McCabe CH, Braunwald E. Effect of Ranolazine, an Antianginal Agent With Novel Electrophysiological Properties, on the Incidence of Arrhythmias in Patients With Non ST-Segment Elevation Acute Coronary Syndrome: Results From the Metabolic Efficiency With Ranolazine for Less Ischemia in Non ST-Elevation Acute Coronary Syndrome Thrombolysis in Myocardial Infarction 36 (MERLIN-TIMI 36) Randomized Controlled Trial. *Circulation.* 2007;116:1647-1652.

7. Spinelli W, Parsons RW, Colatsky TJ. Effects of WAY-123,398, a new Class-III antiarrhythmic agent, on cardiac refractoriness and ventricular fibrillation threshold in anesthetized dogs - a comparison with UK-68798, e-4031, and DL- Sotalol. *J Cardiovasc Pharmacol.* 1992;20:913-922.

8. Wiesfeld AC, De Langen CD, Crijns HJ, Bel KJ, Hillege HL, Wesseling H, Lie KI. Rate-dependent effects of the class III antiarrhythmic drug almokalant on refractoriness in the pig. *J Cardiovasc Pharmacol.* 1996;27:594-600.

9. Baskin EP, Lynch JJ, Jr. Differential atrial versus ventricular activities of class III potassium channel blockers. *J Pharmacol Exp Ther.* 1998;285:135-142.

10. Wang J, Feng J, Nattel S. Class III antiarrhythmic drug action in experimental atrial fibrillation. Differences in reverse use dependence and effectiveness between d-sotalol and the new antiarrhythmic drug ambasilide. *Circulation.* 1994;90:2032-2040.

11. Echt DS, Berte LE, Clusin WT, Samuelson RG, Harrison DC, Mason JW. Prolongation of the human monophasic action potential by sotalol. *Am J Cardiol.* 1982;50:1082-1086.

12. Stump GL, Wallace AA, Regan CP, Lynch JJ, Jr. In vivo antiarrhythmic and cardiac electrophysiologic effects of a novel diphenylphosphine oxide IKur blocker (2-isopropyl-5-methyl-cyclohexyl) diphenylphosphine oxide. *J Pharmacol Exp Ther.* 2005;315:1362-1367.

13. Buchanan LV, LeMay RJ, Walters RR, Hsu CY, Brunden MN, Gibson JK. Antiarrhythmic and electrophysiologic effects of intravenous ibutilide and sotalol in the canine sterile pericarditis model. *J Cardiovasc Electrophysiol.* 1996;7:113-119.

14. Bettoni M, Zimmermann M. Autonomic tone variations before the onset of paroxysmal atrial fibrillation. *Circulation.* 2002;105:2753-2759.

15. Pappone C, Santinelli V, Manguso F, Vicedomini G, Gugliotta F, Augello G, Mazzone P, Tortorello V, Landoni G, Zangrillo A, Lang C, Tomita T, Mesas C, Mastella E, Alfieri O. Pulmonary vein denervation enhances long-term benefit after circumferential ablation for paroxysmal atrial fibrillation. *Circulation.*

2004;109:327-334.

16. Bosch RF, Zeng X, Grammer JB, Popovic K, Mewis C, Kuhlkamp V. Ionic mechanisms of electrical remodeling in human atrial fibrillation. *Cardiovasc Res.* 1999;44:121-131.
17. Ehrlich JR, Cha TJ, Zhang L, Chartier D, Villeneuve L, Hebert TE, Nattel S. Characterization of a hyperpolarization-activated time-dependent potassium current in canine cardiomyocytes from pulmonary vein myocardial sleeves and left atrium. *J Physiol.* 2004;557:583-597.
18. Dobrev D, Friedrich A, Voigt N, Jost N, Wettwer E, Christ T, Knaut M, Ravens U. The G Protein-Gated Potassium Current IK_{ACh} Is Constitutively Active in Patients With Chronic Atrial Fibrillation. *Circulation.* 2005;112:3697-3706.
19. Voigt N, Friedrich A, Bock M, Wettwer E, Christ T, Knaut M, Strasser RH, Ravens U, Dobrev D. Differential phosphorylation-dependent regulation of constitutively active and muscarinic receptor-activated IK_{ACh} channels in patients with chronic atrial fibrillation. *Cardiovasc Res.* 2007;74:426-437.
20. Cha TJ, Ehrlich JR, Chartier D, Qi XY, Xiao L, Nattel S. Kir3-based inward rectifier potassium current: potential role in atrial tachycardia remodeling effects on atrial repolarization and arrhythmias. *Circulation.* 2006;113:1730-1737.
21. Hashimoto N, Yamashita T, Tsuruzoe N. Tertiapin, a selective IK_{ACh} blocker, terminates atrial fibrillation with selective atrial effective refractory period prolongation. *Pharmacol Res.* 2006;54:136-141.
22. Cummings JE, Gill I, Akhrass R, Dery M, Biblo LA, Quan KJ. Preservation of the anterior fat pad paradoxically decreases the incidence of postoperative atrial fibrillation in humans. *J Am Coll Cardiol.* 2004;43:994-1000.
23. Wettwer E, Hala O, Christ T, Heubach JF, Dobrev D, Knaut M, Varro A, Ravens U. Role of IK_r in controlling action potential shape and contractility in the human atrium: influence of chronic atrial fibrillation. *Circulation.* 2004;110:2299-2306.
24. Burashnikov A, Mannava S, Antzelevitch C. Transmembrane action potential heterogeneity in the canine isolated arterially-perfused atrium: effect of IK_r and Ito/IK_{ur} block. *Am J Physiol.* 2004;286:H2393-H2400.
25. Burashnikov A, Antzelevitch C. Can inhibition of IK_{ur} promote atrial fibrillation? *Heart Rhythm.* 2008 (in press).
26. Olson TM, Alekseev AE, Liu XK, Park SJ, Zingman LV, Biengraeber M, Sattiraju S, Ballew JD, Jahangir A, Terzic A. Kv1.5 channelopathy due to KCNA5 loss-of-function mutation causes human atrial fibrillation. *Hum Mol Genet.* 2006;15:2185-2191.
27. Blaauw Y, Gogelein H, Tieleman RG, van HA, Schotten U, Allessie MA. "Early" class III drugs for the treatment of atrial fibrillation: efficacy and atrial selectivity of AVE0118 in remodeled atria of the goat. *Circulation.* 2004;110:1717-1724.
28. Knobloch K, Brendel J, Rosenstein B, Bleich M, Busch AE, Wirth KJ. Atrial-selective antiarrhythmic actions of novel I_{Kur} vs. I_{Kr}, I_{Ks}, and IK_{ACh} class Ic drugs and beta blockers in pigs. *Med Sci Monit.* 2004;10:BR221-BR228.
29. Wirth KJ, Brendel J, Steinmeyer K, Linz DK, Rutten H, Gogelein H. In vitro and in vivo effects of the atrial selective antiarrhythmic compound AVE1231. *J Cardiovasc Pharmacol.* 2007;49:197-206.
30. Regan CP, Stump GL, Wallace AA, Anderson KD, McIntyre CJ, Liverton NJ, Lynch JJ, Jr. In vivo cardiac electrophysiologic and antiarrhythmic effects of an isoquinoline IK_{ur} blocker, ISQ-1, in rat, dog, and nonhuman primate. *J Cardiovasc Pharmacol.* 2007;49:236-245.
31. Dorian P, Pinter A, Mangat I, Korley V, Cvitkovic SS, Beach GN. The effect of vernakalant (RSD1235), an investigational antiarrhythmic agent, on atrial electrophysiology in humans. *J Cardiovasc Pharmacol.* 2007;50:35-40.
32. Goldstein RN, Khrestian C, Carlsson L, Waldo AL. Azd7009: a new antiarrhythmic drug with predominant effects on the atria effectively terminates and prevents reinduction of atrial fibrillation and flutter in the sterile pericarditis model. *J Cardiovasc Electrophysiol.* 2004;15:1444-1450.
33. Seki A, Hagiwara N, Kasanuki H. Effects of NIP-141 on K currents in human atrial myocytes. *J Cardiovasc Pharmacol.* 2002;39:29-38.
34. Matsuda T, Takeda K, Ito M, Yamagishi R, Tamura M, Nakamura H, Tsuruoka N, Saito T, Masumiya H, Suzuki T, Iida-Tanaka N, Itokawa-Matsuda M, Yamashita T, Tsuruzoe N, Tanaka H, Shigenobu K. Atria selective prolongation by NIP-142, an antiarrhythmic agent, of refractory period and action potential duration in guinea pig myocardium. *J Pharmacol Sci.* 2005;98:33-40.
35. Burashnikov A, Di Diego JM, Zygmunt AC, Belardinelli L, Antzelevitch C. Atrial-selective sodium channel block as a strategy for suppression of atrial fibrillation. *Ann NY Acad Sci.* 2008;1123:105-112.
36. Burashnikov A, Antzelevitch C. Atrial-selective sodium channel blockers. Do they exist? *J Cardiovasc Pharmacol.* 2008 (In press).
37. Carlsson L, Chartier D, Nattel S. Characterization of the in vivo and in vitro electrophysiological effects of the novel antiarrhythmic agent AZD7009 in atrial and ventricular tissue of the dog. *J Cardiovasc Pharmacol.* 2006;47:123-132.
38. Fedida D. Vernakalant (RSD1235): a novel, atrial-selective antifibrillatory agent. *Expert Opin Investig Drugs.* 2007;16:519-532.
39. Regan CP, Kiss L, Stump GL, McIntyre CJ, Beshore DC, Liverton NJ, Dinsmore CJ, Lynch JJ, Jr. Atrial antifibrillatory effects of structurally distinct IK_{ur} blockers 3-[(dimethylamino)methyl]-6-methoxy-2-methyl-4-phenylisoquinolin-1(2H)-one and 2-phenyl-1,1-dipyridin-3-yl-2-pyrrolidin-1-yl-ethanol in dogs with underlying heart failure. *J Pharmacol Exp Ther.* 2008;324:322-330.
40. Wirth KJ, Paehler T, Rosenstein B, Knobloch K, Maier T, Frenzel J, Brendel J, Busch AE, Bleich M. Atrial effects of the novel K(+)-channel-blocker AVE0118 in anesthetized pigs. *Cardiovasc Res.* 2003;60:298-306.
41. Antzelevitch C, Shimizu W, Yan GX, Sicouri S, Weissenburger J, Nesterenko VV, Burashnikov A, Di Diego JM, Saffitz J, Thomas GP. The M cell: its contribution to the ECG and to normal and abnormal electrical function of the heart. *J Cardiovasc Electrophysiol.* 1999;10:1124-1152.
42. Burashnikov A, Antzelevitch C. Late-Phase 3 EAD. A Unique Mechanism Contributing to Initiation of Atrial Fibrillation. *PACE.* 2006;29:290-295.
43. Amos GJ, Wettwer E, Metzger F, Li Q, Himmel HM, Ravens U. Differences between outward currents of human atrial and subepicardial ventricular myocytes. *J Physiol.* 1996;491 (Pt 1):31-50.
44. Nattel S, Matthews C, De Blasio E, Han W, Li D, Yue L. Dose-dependence of 4-aminopyridine plasma concentrations and electrophysiological effects in dogs: potential relevance to ionic mechanisms in vivo. *Circulation.* 2000;101:1179-1184.
45. Burashnikov A, Belardinelli L, Antzelevitch C. Ranolazine and propafenone both suppress atrial fibrillation but ranolazine unlike propafenone does it without prominent effects on ventricular myocardium. *Heart Rhythm.* 2007;4:S163 (Abstract).
46. Li GR, Lau CP, Shrier A. Heterogeneity of sodium current in atrial vs epicardial ventricular myocytes of adult guinea pig hearts. *J Mol Cell Cardiol.* 2002;34:1185-1194.
47. Whalley DW, Wendt DJ, Grant AO. Basic concepts in cellular cardiac electrophysiology: Part II: Block of ion channels

by antiarrhythmic drugs. *PACE*. 1995;18:1686-1704.

48. Crijns HJ, Van G, I, Walfridsson H, Kulakowski P, Ronaszeki A, Dedek V, Malm A, Almgren O. Safe and effective conversion of persistent atrial fibrillation to sinus rhythm by intravenous AZD7009. *Heart Rhythm*. 2006;3:1321-1331.

49. Roy D, Pratt CM, Torp-Pedersen C, Wyse DG, Toft E, Juul-Moller S, Nielsen T, Rasmussen SL, Stiell IG, Coutu B, Ip JH, Pritchett EL, Camm AJ. Vernakalant Hydrochloride for Rapid Conversion of Atrial Fibrillation. A Phase 3, Randomized, Placebo-Controlled Trial. *Circulation*. 2008;117:1518-1525.

50. Fuster V, Ryden LE, Cannom DS, Crijns HJ, Curtis AB, Ellenbogen KA, Halperin JL, Le Heuzey JY, Kay GN, Lowe JE, Olsson SB, Prystowsky EN, Tamargo JL, Wann S, Smith SC, Jr., Jacobs AK, Adams CD, Anderson JL, Antman EM, Hunt SA, Nishimura R, Ornato JP, Page RL, Riegel B, Priori SG, Blanc JJ, Budaj A, Camm AJ, Dean V, Deckers JW, Despres C, Dickstein K,

Lekakis J, McGregor K, Metra M, Morais J, Osterspey A, Zamorano JL. ACC/AHA/ESC 2006 guidelines for the management of patients with atrial fibrillation--executive summary: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines and the European Society of Cardiology Committee for Practice Guidelines (Writing Committee to Revise the 2001 Guidelines for the Management of Patients With Atrial Fibrillation). *J Am Coll Cardiol*. 2006;48:854-906.

51. Duytschaever M, Blaauw Y, Allessie M. Consequences of atrial electrical remodeling for the anti-arrhythmic action of class IC and class III drugs. *Cardiovasc Res*. 2005;67:69-76.

52. Linz DK, Afkham F, Itter G, Rutten H, Wirth KJ. Effect of Atrial Electrical Remodeling on the Efficacy of Antiarrhythmic Drugs: Comparison of Amiodarone with I(Kr)- and I(to)/IKur-Blockade In Vivo Atrial Electrical Remodeling and Antiarrhythmic Drugs. *J Cardiovasc Electrophysiol*. 2007;18:1313-1320.